

PROTEASES OF THE SEEDS OF THE COTTON PLANT

II. SEPARATION AND PURIFICATION

G. M. Podgornov, M. A. Kuchenkova,
and P. Kh. Yuldashev

UDC 577.156

We separated the fractions with protease activity [1] and purified them by gel filtration on a column of Sephadex G-100 (column 4.5×100 cm; rate of elution 60 ml/h) using 0.1 M phosphate buffer with pH 7.4. Two active fractions were obtained: 1 and 2. Fraction 1 was then passed through a column of Sephadex G-200 (column 2×75 cm, rate of elution 7 ml/h, 0.1 M phosphate buffer, pH 7.4). The active fractions obtained were called protease I and protease II. Fraction 2 after gel filtration through G-100 was passed through a column of DEAE-cellulose equilibrated with 0.1 M phosphate buffer, pH 7.4 (column 1×20 cm, rate of elution 16 ml/h). The protein was eluted by the imposition of a gradient: 0.01 M phosphate buffer with pH 7.4-0.1 M phosphate buffer with pH 7.4 and 0.5 M NaCl. Two active fractions were obtained: one before the imposition of the gradient and the other after it. The fraction eluted after the imposition of the gradient at a concentration of NaCl of 0.1-0.15 M proved to have the maximum specific activity and was called protease III. Experimental information on the separation and purification processes is given in Table 1.

The disk electrophoresis in polyacrylamide gel [2] of proteases I and II showed a single clear band for each and two very weak brightenings while in the case of protease III there was one broad band and five weak brightenings. The molecular weights of proteases I-III were determined by gel filtration through a column of Sephadex G-200; they were, respectively, 140,000, 70,000, and 30,000. As markers for calibration (column 2×75 cm, rate of elution 7 ml/h) we used "Reakhim" catalase (mol. wt. 240,000), "Reanal" lactate dehydrogenase from porcine cardiac muscle (mol. wt. 142,000), "Hungary" bovine serum albumin (mol. wt. 67,000), and "Spofa" trypsin (mol. wt. 24,000). The free volume of the column was determined with respect to dextran blue "Pharmacia."

TABLE 1

Stage of purification	Protein content, g	Proteolytic activity		Yield, % with respect to		Degree of purification
		specific	total	protein	activity	
First extract in 0.1 M phosphate buffer, pH 7.4	5.2	20	104	—	—	—
Fraction 0-60% saturation $[(\text{NH}_4)_2\text{SO}_4]$, separation on Sephadex G-100						
Fraction 1	1.26	70	88.2	24.2	80	3.5
Fraction 2	0.95	40	38	18.6	36.5	2
Separation on Sephadex G-200 of Fraction 1 after G-100	0.032	430	13.7	0.6	13.5	21
1 (protease I)	0.84	50	42	16.2	41	2.5
2 (protease II)	0.13	170	22.1	2.4	21.2	8.5
Separation on DEAE-cellulose of Fraction 2 after G-100	0.037	530	19.6	0.7	19.3	26.5
1	0.032	430	13.7	0.6	13.5	21
2 (protease III)	0.016	278	4.98	0.31	5	13
	0.008	800	6.4	0.11	6.1	40

Order of the Red Banner of Labor Institute of the Chemistry of Plant Substances, Academy of Sciences of the Uzbek SSR. Translated from *Khimiya Prirodnikh Soedinenii*, No. 1, pp. 120-121, January-February, 1974. Original article submitted June 5, 1973.

© 1975 Plenum Publishing Corporation, 227 West 17th Street, New York, N.Y. 10011. No part of this publication may be reproduced, stored in a retrieval system, or transmitted, in any form or by any means, electronic, mechanical, photocopying, microfilming, recording or otherwise, without written permission of the publisher. A copy of this article is available from the publisher for \$15.00.

Thus, three forms of proteases with dissimilar molecular weights have been found in dormant cotton seeds and a method for their separation has been developed.

LITERATURE CITED

1. M. A. Kuchenkova, N. L. Ovchinnikova, and P. Kh. Yuldashev, *Khim. Prirodn. Soedin.*, 4 (1973).
2. K. Davranov, M. A. Kuchenkova, and P. Kh. Yuldashev, *Khim. Prirodn. Soedin.*, 806 (1971).